

#sharing challenges and solutions in practice

Review of Online water monitoring analyzers (OWBA) and their potential application

GMP/FDA Compliance Conference Dr. Hans-Joachim Anders, Novartis Pharma

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Dr. Hans-Joachim Anders

University Ulm

- Study of biology with focus on microbiology and molecular biology
- PhD in microbiology on the anaerobic metabolism of aromatic compounds in denitrifying bacteria.
- MSD (Merck, Sharp & Dohme) and Fresenius Home Care
 - Field service sales and patient care
- Novartis Pharma AG, Stein, Aargau, Switzerland
 - Lab Head, Senior QA Facilitator, Teamlead Analytical Science & Technology Microbiology, microbiological quality assurance and control



 Member of the expert group of the European Pharmacopoeia for pharmaceutical water







Agenda

- Online Water Monitoring technologies
 - Biofluorescent particle counters
 - Flow Cytometry
 - Validation aspects
- Evaluation study Flow Cytometry
 - Result interpretation higher counts and possible explenation
- Potential use of Online Water Monitoring Technologies
- Views of the industry working group collaboration (BPHOG/OWBA)
 - Who we are?
 - Expected benefits and vision of the future
 - Why isn't it reality already?









Online detection methods

- Continuous monitoring.
- Direct connection of the analyzer to the pharmaceutical water treatment or distribution system at line.
- No laboratory analysis necessary.
- Biofluorescent Particle Counter
 - ⇒ Detection of the microorganism by excitation with a laser.
 - ⇒ Instruments measure the auto-fluorescence of the microorganisms.
- Flow Cytometry
 - ⇒ Viability stain
 - ⇒ Detection of stained viable cells







Examples manufacturer and instruments



Mettler-Toledo RMS7000 https://www.mt.com/



Sentinel MOBA http://www.sentinelmonitors.com/



AQU@Sense MB https://www.bwt-pharma.com/

Biofluorescent particle counters







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Biofluorescent particle counter

• Detection principle



RMS 7000 Flow Cell









Biofluorescent particle counter

Similarly to a Total Particle Counter...



Each particle is assessed for: Scattered light intensity Fluorescent light intensity

Continuous, real-time outputs: Total particle counts Auto-fluorescence counts (AFU)







Biofluorescent particle counter

- Detection principle
 - ⇒ Excitation with 405 nm
 - ⇒ Autofluorescence of ATP, NAD(P)H and Riboflavine



- "Real-time Measurement of Fluorescence Spectra from Single Airborne Biological Particles", Hill et al., Field Analytical Chemistry and Technology, 3, 221–239 (1999) (265 nm excitation)
- "High performance recycling of polymers by means of their fluorescence liftetimes". Heinz Langhals et al. Dept of Chemistry, LUM University of Munich, Munich, Germany. August 2014 (365 nm excitation)

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Information Source

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- Biofluorescent particle counter
 - Result in Autofluorescence Units AFU





- Colony-forming unit (CFU) is a unit that gives an estimate of the number of microorganisms in a sample.
- Auto-Flourescence Unit (AFU) is a unit that takes into account fluorescence as well as size of a particle.





Biofluorescent particle counter

- Points to consider:
- Qualification aspects:
 - ⇒ False-positive rate
 - ⇒ Calibration standard
 - Calibration
 - Suitability
- Validation aspects



9 days of WFI monitoring







Biofluorescent particle counter

- Reason for False-positive signals
 - ⇒ Microorganisms in the VNBC stage, Viable but not culturable, described for pharmaceutical water.
 - ⇒ Microorganisms that do not grow on the media used but can be detected during online measurement (95-99% of all microorganism species or have not been cultivated with traditional methods so far, e.g. because the media/conditions used do not allow growth).
 - ⇒ Polymers (EPDM, Teflon), dead cells, pollen, some solvents (isopropanol), rouge, metal abrasion (reflective properties) can lead to false positive results.







Biofluorescent particle counter

- Validation aspects:
- AFU \neq CFU and

rate of false-positive results and their origin

- ⇒ Will make comparative studies using the traditional method difficult or impossible.
- ➡ Most test criteria to validate an alternative microbiological method according to EP 5.1.6, USP<1223> or PDA TR33 can't be tested
 - especially equivalence to traditional method.
 - Spiking experiments are of limited significance due to particle contamination and viability stage of the organisms compared to organisms in pharmaceutical grade water
- ⇒ Possible solutions
 - Signal induced auto sampler will allow traditional anlysis of the peak
 - Alternative assessment of equivalence according to USP1223 Decision Equivalence.







Biofluorescent particle counter

- Calibration of the instrument
 - ⇒ No model organism available
 - Vitality and size of the microorganisms and thus the fluorescence signal depends on the growth phase in which the microorganisms are located, i.e. an organism cultured on medium is not comparable with a microorganism adapted to pharmaceutical water.
 - ⇒ OWBA working group is therefore pursuing the establishment of fluorescent beads similar in size to Ralstonia pickettii.
 - ⇒ Ralstonia pickettii is a gram-negative rod-shaped bacterium that can often be isolated from pharmaceutical water.



http://www.higieneambiental.com/calidad-de-aireinterior/ralstonia-pickettii-patogeno-oportunista-emergente







Biofluorescent particle counter

- Calibration of the instrument
 - ⇒ In collaboration with NIST (National Institute of Standards and Technology), a replacement microorganism in the form of fluorescent beads will be developed.
 - ⇒ Like online TOC with artificial standards sucrose/benzoquinone.
 - ⇒ The following properties must be met by the standard:
 - Number
 - Size
 - Fluorescence (critical parameter, because the fluorescence of commercial beads are too high)
 - Stability
 - ⇒ Comparative studies with Ralstonia pickettii.







- Flow Cytometry Aqu@Sense
 - Overview
 - Automated Flow Cytometry
 - Continuous monitoring of microorganisms in pharmaceutical grade water
 - Measurement interval from 0,5 6 h
 - Stand alone (Offline) or integrated use (Online) possible
 - Instrument sanitizable with Ozone or hot water











- Flow Cytometry
 - Viability stain









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Flow Cytometry

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- Validation aspects:
 - ⇒ Instrument/Method could be validated according EP 5.1.6, USP<1223> or PDA TR33.
 - ⇒ Cell count vs. CFU
 - ⇒ Based on viability staining, interfering particles probably play a lesser role.
 - ⇒ Offline mode will allow to use discrete samples for validation testing.
 - ⇒ Spiking and equivalence tests are possible.







Flow Cytometry – Feasibility Study

- Feasibility Study Tests:
 - ⇒ Accuracy and Precision with P. aeruginosa, S. maltophilia, S. aureus, M. radiotolerans
 - ⇒ Equivalence «real» samples compared to plate count
- Additional tests:
 - ⇒ Linearitiy different dilutions of microorganisms
 - ⇒ Background
 - ⇒ Detection of Spore-forming microorganism







- Flow Cytometry Feasibility Study results
 - Accuracy/Precision

Stenotrophomonas maltophilia

Accuracy	per mL (90 µL analyzed)	AquaSense replicate 1 AquaSense replicate 2	1978 1866	1974 1855	2134 2172
The mean of Aqu@Sense values for <i>S. maltophilia</i> is significantly greater.		AquaSense replicate 3 AquaSense replicate 4 AquaSense replicate 5	1918 1808 2035	2004 1888 2166	2326 2825 2264
Precision	per mL (90 µL plated)	Pour plate replicate 1 Pour plate replicate 2 Pour plate replicate 3	1622 1256 1322	1611 1722 1611	1611 1789 2178
Stdevs are NOT significantly different		Pour plate replicate 4 Pour plate replicate 5	1311 1344	1467 1356	1511 1633





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Run 2

Run 1

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Run 3

- Flow Cytometry Feasibility Study results
 - Accuracy/Precision

Methylobacterium radiotolerans

<u>Accuracy</u>	per mL (90 µL analyzed)	AquaSense replicate 1 AquaSense replicate 2	1807 2079	769 922	474 571
The mean of Aqu@Sense values for <i>M. radiotolerans</i> is NOT significantly greater.		AquaSense replicate 3 AquaSense replicate 4 AquaSense replicate 5	2017 2028 2138	971 922 761	675 748 761
Precision	per mL (90 µL plated)	Pour plate replicate 1 Pour plate replicate 2 Pour plate replicate 3	2489 2667 2589	1111 1033 1211	767 767 700
Stdevs are NOT significantly different		Pour plate replicate 4 Pour plate replicate 5	2756 2456	1144 1067	656 711





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Run 2

Run 1

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Run 3

- Flow Cytometry Feasibility Study results
 - Accuracy/Precision

Pseudomonas aeruginosa Run 1 Run 2 Run 3 458 per mL AquaSense replicate 1 441 571 Accuracy AquaSense replicate 2 (90 µL analyzed) 886 740 495 AquaSense replicate 3 852 842 709 The mean of Aqu@Sense values for AquaSense replicate 4 969 731 776 P. aeruginosa is significantly less. AquaSense replicate 5 1006 970 943 per mL Pour plate replicate 1 789 911 1144 Precision (90 µL plated) Pour plate replicate 2 1067 933 1278 Pour plate replicate 3 856 722 1156 Pour plate replicate 4 856 1067 1144 Stdevs are NOT significantly different Pour plate replicate 5 1078 889 1033







- Flow Cytometry Feasibility Study results
 - Equivalence

After Reverse Osmosis stage 1

		Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10	Day 11
AquaSense replicate 1 AquaSense replicate 2	per ml	66 54	76 99	131 110	76 67	43 54	121 123	77 65	76 99	109 98	101 143	109 88
Pour plate replicate 1 Pour plate replicate 2	per ml	11 11	0 0	56 22	22 11	0 22	44 78	44 11	11 22	0 0	16 23	20 17



Test: Mean(115_AS_all_day1-11) / Mean(115_PP_all_day1-11) > Lower Limit

95% Lower bound for Mean(115_AS_all_day1-11) / Mean(115_PP_all_day1-11): 3.2400 Lower bound is greater than 0.8. Can claim Mean(115_AS_all_day1-11) / Mean(115_PP_all_day1-11) > 0.8





- Flow Cytometry Feasibility Study results
 - Equivalence

After Softener

		Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10	Day 11
AquaSense replicate 1 AquaSense replicate 2	per ml	5038 5490	3677 3624	6164 6622	4985 5083	3327 3514	4194 3666	2955 2596	3041 3377	3275 3674	3068 3123	4677 4477
Pour plate replicate 1 Pour plate replicate 2	per ml	167 278	1256 1500	933 833	388 411	289 356	244 289	178 189	922 700	667 544	389 389	511 389





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Flow Cytometry – Feasibility Study results

• Why higher counts in Equivalence studies?

Two water samples from two sampling points after Softener, were tested in duplicates with the ScanRDI, Aqu@Sense and Pour plate method.

ScanRDI: 1 ml of each sample was filtrated with 9 ml of sterilized Purified Water and then processed according the direct detection protocol.

Aqu@Sense: $2 \times 90 \mu$ L of each water sample are analyzed automatically by the Aqu@Sense BW. Result is given per mL!

Pour plate: 2 x 90 μ L and 2 x 1 mL of each water sample are pipetted in an empty petri dish and covered with approximately 20 mL R2A Agar, 30-35°C, 5 – 7 days







- Flow Cytometry Feasibility Study results
 - Why higher counts in Equivalence studies?

Method	Sampling Point 1 Sample1	Sampling Point 1 Sample 2	Sampling Point 2 Sample1	Sampling Point 2 Sample 2
ScanRDI (events/MO per ml)	46	43	115	107
Aqu@Sense (ICC/ml)	1403	1453	2084	1572
Pour plate (cfu/ml)	0	4	38	23







- Flow Cytometry Feasibility Study results
 - Why higher counts in Equivalence studies?





- Flow Cytometry Feasibility Study results
 - Why higher counts in Equivalence studies?









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- Flow Cytometry Feasibility Study results
 - Why higher counts in Equivalence studies? Diagrams





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- Flow Cytometry Feasibility Study results
 - Why higher counts in Equivalence studies? Conclusion
 - ⇒ The cell count measurement with the Aqu@Sense method for samples after Softener is approx. 32/16 times higher than with the ScanRDI method.
 - ⇒ Viable but not culturable cells with Scan RDI the could be detected, could explain the lower plate count results.
 - Plate Counts and ScanRDI counts lower due to clustering of organisms, as can be seen in the ScanRDI pictures.







Both Technologies

- Potential use of the Online instruments in the view of OWBA workgroup:
 - ⇒ Optimization of processes like sanitisation
 - ⇒ Revalidation after maintenance work, Pre- und Post-Maintenance measurement
 - ⇒ Biofilm-Monitoring
 - ⇒ Reducing of grab sampling in water treatment plants
 - Publication: Anders HJ, Ayers F, Fitch B, Forng RY, Hooper S, Luebke M, Mateffy J, Noverini P, Termine B, Yan L, and Weber J of the Online Water Bioburden Analyzer Workgroup(2017): Practical Application Of Online Water Bioburden Analyzers In Pharmaceutical Manufacturing

https://www.pharmaceuticalonline.com/doc/practical-application-of-online-water-bioburden-analyzers-in-pharmaceutical-manufacturing-0001?vm_tld=2016150&user=f7b92eb0-a95c-4087-a7dd-dfd35afdbc01&utm_source=et_6214180&utm_medium=email&utm_campaign=PHARM_08-15-2017&utm_term=f7b92eb0-a95c-4087-a7dddfd35afdbc01&utm_content=Practical+Application+Of+Online+Water+Bioburden+Analyzers+In+Pharmaceutical+Manufacturing_







Both Technologies

Pro's	Con's
Online Analysis	Cell count, AFU ≠ CFU
Continuous Monitoring	Authority acceptance?
No lab analysis, no dependency on media properties	Alternative method, Validiation acc. EP 5.1.6, USP<1223>, PDA TR33 ? Equivalence?
Sensitive Detection with Laser, Autofluorescence, Viability stain (no growth necessary)	Calibration
Non destructive method, Auto sampler?	False-positive signals of particles (Kunststoffe, etc.), VNBC
Data Integrity	Investment costs



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 Industry working group established to discuss and push the implementation of Biofluorecent Particle counter

Who we are:

Philip Villari, Merck & Co., Inc. (Author of the following slides) Joanny Salvas, Pfizer (Author of the following slides) Hans-Joachim Anders, Novartis Pharma Stein AG James Cannon, Mettler-Toledo Thornton, Inc. Anthony Cundell, Microbiological Consulting, LLC. Michael Dingle, TSI Inc. David Govezensky, Bio-Technology General (Israel) Ltd. Patrick Hutchins, TSI Inc. Cedric Joossen, Janssen Chris Knutsen, Bristol-Myers Squibb Petra Merker, Bayer AG Stephanie Ramsey, Amgen Inc. Margit Franz-Riethdorf, BioPhorum Allison Scott, ANAD BioVigilant Ans Vanbroekhoven, Sanofi











Disclaimer

The information and opinions presented are those of the collaboration, and not necessarily the opinions of our individual employers.







Who we are – Members:









Publications:

Paper 1: Challenges encountered during BFPC implementation

Understanding the non-equivalency of AFUs and CFUs

> Proposal for validation strategy of BFPC for use for GMP decisions

Proposal for establishing alert and action limits for BFPC used in GMP areas

. TBD

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Expected benefits and vision of the future

Improved Product Quality Control

Real Time EM Data

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- Reduced Manufacturing Risks
- Improved Root Cause Analysis
- Process Understanding





Enhances the Marketing and Supply of Safer and Effective Medicines to Patient Population









Expected benefits and vision of the future



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Why aren't Online Monitoring instruments reality already?

Colony-forming unit (CFU) is a unit used to estimate the number of viable and culturable bacteria or fungal cells in a sample

Auto-Fluorescence Unit (AFU) is a unit that reflects both size and fluorescence of the particle that can detect viable but non-culturable cells in a sample









- Why aren't Online Monitoring instruments reality already?
 - What can be measured as AFUs and/or CFUs?



Relative sizes are for illustration purposes only and may differ from sample to sample







Why aren't Online Monitoring instruments reality already?



•







- Why aren't Online Monitoring instruments reality already?
- Validation challenges:
 - Why is validation a challenge?

⇒Require different methodology than traditional method (AFU ≠ CFU)
 ⇒Interpretation of validation guidance can be difficult
 ⇒Extensive validation is often expected

• Overcoming this challenge

⇒Utilize Industry Working Group and Regulatory Support

⇒Determine extent of validation appropriate to application

⇒FDA safe harbor principle and research exemption use as technology is evaluated and implemented

Additional Information

⇒M³ collaboration has a paper on validation in progress which will provide a detailed approach







- Why aren't Online Monitoring instruments reality already?
- Validation strategy:
 - Primary vendor validation address Limit of Detection, Limit of Quantification, etc.
 - User establish non-inferiority to traditional method
 - USP <1223> offers four validation options regarding equivalence testing:
 - ⇒ Decision equivalence: most applicable
 - Compare ability of BFPC vs. Traditional to identify out-of-limit event
 - M³ Working Group for validation strategy paper
- Qualification Strategy
 - BFPC testing in desired environment to establish baseline counts and potential interferents; "Tailor BFPCs to your needs!"
 - In situ testing in Grades C and D areas or water treatment plants (non-zero AFU and CFU counts)
 - ⇒ Obtain natural microflora information







- Why aren't Online Monitoring instruments reality already?
- Setting Alert and Action Levels
 - AFU determination often more sensitive than CFU
 - Continuous monitoring can overestimate meaningful excursions
 - Difficult to determine sufficient side-by-side testing needed with traditional method







- Why aren't Online Monitoring instruments reality already?
- Setting Alert and Action Levels



M³ collaboration has baseline paper to overcome challenges in progress







Dilemma?

Regulatory guidelines, such as the EU Annex I draft, have been encouraging the use of alternative technologies.





However, these same guidance documents continue to include limits defined in terms of CFU counts, and agencies continue to expect conventional capture and identification of any "hits" or AFUs.

BFPC and their mode of detection of microorganisms require a rethinking across the pharmaceutical industry for manufacturers, inspectors, and regulators to enable a paradigm shift from the traditional to modern monitoring methods.









Thank you for your attention

QUESTIONS?











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